

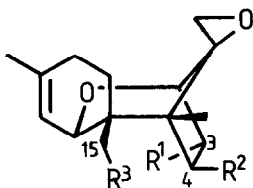
CONVERSION OF ANGUIDINE INTO CALONECTRIN AND 3-DEACETYL-CALONECTRIN <sup>1</sup>

Nicolas Jeker, Peter Mohr & Christoph Tamm\*

Institute of Organic Chemistry, University of Basel,  
St. Johannis-Ring 19, CH-4056 Basel, Switzerland

ABSTRACT: Anguidine (diacetoxyscirpenol, 2) was converted into calonectrin (3) in 7 steps using the Barton deoxygenation as key reaction.

The trichothecenes are a class of natural products produced by moulds, especially various species of Fungi imperfecti. Many members of the family display a wide range of biological activities <sup>3</sup>. Both the simple sesquiterpenes and their macrocyclic di- and triesters have attracted much attention from the synthetic, biosynthetic and pharmacological view point. It seems now well established that not only the sesquiterpene moiety in the naturally occurring metabolites, most often verrucarol (1), but also the polyfunctionalized macrocyclic portion is relevant for the biological activity. In connection with our interest in preparing novel, unnatural macrocycles, we studied the functional group interchange of anguidine (diacetoxyscirpenol, 2) <sup>4</sup>. In this communication we report a relatively efficient conversion of this metabolite readily available from various Fusarium strains into 3-deacetyl-calonectrin (3) and calonectrin (4) <sup>5</sup>.



- |          |  |                        |
|----------|--|------------------------|
| <u>1</u> | R <sup>1</sup> =H, R <sup>2</sup> =OH, R <sup>3</sup> =OH  | Verrucarol             |
| <u>2</u> | R <sup>1</sup> =OH, R <sup>2</sup> =R <sup>3</sup> =OAc    | Anguidine              |
| <u>3</u> | R <sup>1</sup> =OH, R <sup>2</sup> =H, R <sup>3</sup> =OAc | 3-Deacetyl-calonectrin |
| <u>4</u> | R <sup>1</sup> =R <sup>3</sup> =OAc, R <sup>2</sup> =H     | Calonectrin            |
| <u>5</u> | R <sup>1</sup> =H, R <sup>2</sup> =OH, R <sup>3</sup> =H   | Trichodermol           |

Our strategy was analogous to that of Tulshian and Fraser-Reid which they followed in the transformation of 2 into verrucarol (1) and trichodermol (5) <sup>6</sup>. Key reaction was a Barton-deoxygenation via the corresponding thiocarbonyl-imidazole derivative <sup>7</sup>. The yield was nearly quantitative. Another crucial step was the chemical differentiation of the C(4)- and C(15)-hydroxyl groups. Whereas silylation (TBDMS-Cl/DMAP/NEt<sub>3</sub>) <sup>8</sup> gave erratic results, acetylation

proceeded with fairly good selectivity comparable to that observed in the verrucarol series <sup>6,9</sup>. The scheme summarizes the reaction sequence leading to 3-deacetyl-calonecetrin (3) and calonecetrin (4) in 6, respectively 7 steps with 30% overall yield for 4. The final products were characterized and identified as follows <sup>10</sup>. Deacetylcalonecetrin (3): m.p. 142.5-148° (hygr.), Lit. 144-145°, microanalysis (found C 65.92%, H 7.92%; calc. C 66.21%, H 7.85%); IR (KBr): 3460 (br), 3060, 3020, 2975, 2945, 1740, 1680, 1245; MS (EI, 10eV): 308 (M<sup>+</sup>, 0.3%), 265 (5.0%), 249 (18%), 248 (100%), 220 (98%); NMR spectra see Table 1 and 2. Calonecetrin (4): m.p. (82.8-85.3°, Lit. 83-85°),  $[\alpha]_D^{27} = +5.8^\circ$  (Lit. +14.6°, authentic reference sample +6.2°, CHCl<sub>3</sub>), microanalysis (found C 64.85%, H 7.44%; calc. C 65.12%, H 7.48%). IR (KBr): 2980, 2940, 2895, 1745, 1730, 1680, 1245. MS (EI, 50eV): 351 (MH<sup>+</sup> 4.26%), 307 (1.3%), 291 (18.5%), 262 (13.5%), 41 (100%); NMR spectra see Table 1 and 2.

**Experimental Procedures:** 3-O-THP,15-O-acetyl-scirpentriol: 2.00 g (5.46 mmol) of diol 6 was dissolved in 40 ml of CH<sub>2</sub>Cl<sub>2</sub> and treated with 2.64 ml of pyridine and 1.03 ml of Ac<sub>2</sub>O (2 eq.). After standing at RT for 14 h, the reaction mixture was diluted with ether, washed with 1N HCl and sat. NaHCO<sub>3</sub>, dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated i.v. column chromatography of the residue (SiO<sub>2</sub>/ether) yielded 1.296 g of the 15-monoacetylated product.

3-Deacetyl-3-O-THP-calonecetrin (7): 201 mg (0.49 mmol) of the above prepared alcohol were refluxed for 6.5 h in 6 ml of 1,2-dichloromethane containing 211 mg (1.19 mmol) of N,N-thiocarbonyldiimidazole. After cooling, CH<sub>2</sub>Cl<sub>2</sub> was added and washed with 1N HCl, sat. NaHCO<sub>3</sub> and brine. The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and evaporated to dryness (253 mg). 238 mg of this crude product were dissolved in 11 ml of toluene and added dropwise during 0.5 h in an Argon atmosphere to 0.243 ml (0.92 mmol) of nBu<sub>3</sub>SnH. After additional 2 h at 110° the reaction mixture was cooled and evaporated i.v. column chromatography of the residue (SiO<sub>2</sub>/ether) yielded 170 mg of 6.

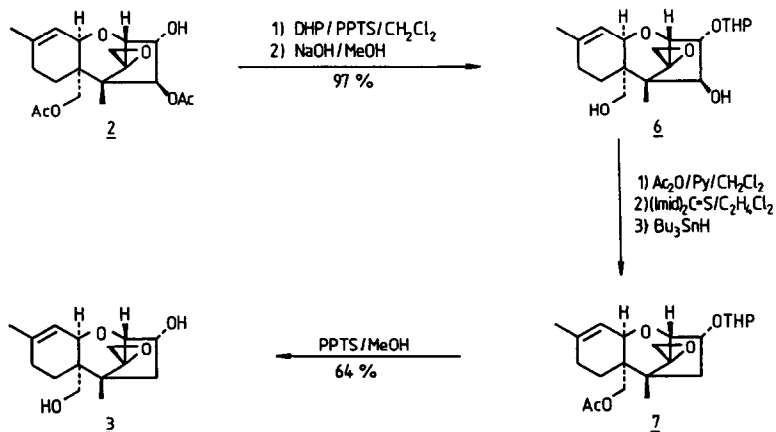


Table 1:  $^1\text{H}$  NMR (200 MHz,  $\text{CDCl}_3$ )

	3-Deacetylcalonecetrin (3)	Calonecetrin (4)
2-H	3.53 (d, $J = 4.5$ , 1 H)	3.76 (d, $J = 5$ , 1 H)
3-H	4.47 (m, 1 H), dx dx d: after exchange with $\text{D}_2\text{O}$	5.17 (dx dx d, 1 H)
4-H	1.5 - 2.3 (m)	1.5 - 2.3 (m)
7-H	1.5 - 2.3 (m)	1.5 - 2.3 (m)
8-H	1.5 - 2.3 (m)	1.5 - 2.3 (m)
10-H	5.51 (d (br), $J = 5$ , 1 H)	5.47 (d (br), $J = 5$ , 1 H)
11-H	4.15 (d (br), $J = 5$ , 1 H)	4.03 (d (br), $J = 5$ , 1 H)
13-H	2.86 and 3.09 (2xd, $J = 4$ , 2 H)	2.86 and 3.10 (2xd, $J=4$ , 2 H)
14-H	0.82 (s, 3 H)	0.83 (s, 3 H)
15-H	3.85 and 4.09 (2xd, $J = 12$ , 2 H)	3.85 and 4.09 (2xd, $J=12$ , 2 H)
16-H	1.73 (s (br), 3 H)	1.73 (s (br), 3 H)
OAc	2.06 (s, 3 H)	2.05 (s, 3 H)
OAc	--	2.12 (s, 3 H)

Table 2:  $^{13}\text{C}$  NMR (22,63 MHz,  $\text{CDCl}_3$ )

	3-Deacetylcalonecetrin (3)	Calonecetrin (4)
C-2	68.4 (d)	68.3 (d)
C-3	69.1 (d)	71.4 (d)
C-4	42.4 (t)	39.6 (t)
C-5	45.9 (s)	45.5 (s)
C-6	43.1 (s)	43.2 (s)
C-7	21.2 (t)	21.0 (t)
C-8	28.4 (t)	28.3 (t)
C-9	140.1 (s)	140.1 (s)
C-10	119.3 (d)	119.2 (d)
C-11	80.0 (d)	78.2 (d)
C-12	65.6 (s)	65.1 (s)
C-13	48.4 (t)	48.5 (t)
C-14	12.3 (q)	12.2 (q)
C-15	62.8 (t)	63.8 (t)
C-16	23.2 (q)	23.1 (q)
$\text{OCOCH}_3$	21.0 (q)	21.2 (2xq)
$\text{OCH}_3$	170.6 (s)	170.5 (s)
		170.2 (s)

ACKNOWLEDGEMENT: We thank the Swiss National Science Foundation for the support of these investigations.

## REFERENCES

- 1 42nd Communication on Verrucarins and Roridins <sup>2</sup>.
- 2 41st Communication: P. Mohr, Ch. Tamm, W. Zürcher & M. Zehnder, *Helv. Chim. Acta* 67, 406 (1984).
- 3 a) Ch. Tamm, *Fortschr. Chem. Org. Naturst.* 31, 63 (1974);  
b) J. R. Bamberg and F. M. Strong in "Microbial Toxins"; S. Kadis, A. Ciegler, C. J. Ajl, Eds.; Academic Press: New York 1973;  
c) T. W. Doyle and W. T. Bradner in "Anticancer Agents Based on Natural Product Models"; I. M. Cassidy and J. Duros, Eds.; Academic Press: New York 1980.
- 4 Isolation and structure of anguidine:  
a) A. W. Dawkins, *J. Chem. Soc. C* 1966, 116;  
b) H. P. Sigg, R. Mauli, E. Flury and D. Hauser, *Helv. Chim. Acta* 48, 962 (1965).
- 5 For unnatural derivatives of anguidine see: T. Kaneko, H. Schmitz, J. M. Essery, W. Rose, H. G. Howell, F. A. O'Herron, S. Nachfolger, J. Huftalen, W. T. Bradner, R. A. Partyka, T. W. Doyle, J. Davies and E. Cundliffe, *J. Med. Chem.* 25, 579 (1982).
- 6 D. B. Tulshian and B. Fraser-Reid, *Tetrahedron Letters* 21, 4549 (1980).
- 7 D. H. R. Barton and S. W. McCombie, *J. Chem. Soc., Perkin Trans. I* 1975, 1574; for a review see: W. Hartwig, *Tetrahedron* 1983, 2609.
- 8 S. K. Chaudhary and O. Hernandez, *Tetrahedron Letters* 20, 99.
- 9 see e.g. a) W. C. Still and H. Ohmizu, *J. Org. Chem.* 46, 5242 (1981);  
b) W. R. Roush and T. A. Blizzard, *J. Org. Chem.* 48, 758 (1983);  
c) P. Mohr, M. Tori, P. Grossen, P. Herold and Ch. Tamm, *Helv. Chim. Acta* 65, 1412 (1982).
- 10 D. Gardner, A. T. Glen and W. B. Turner, *J. Chem. Soc., Perkin Trans. I* 1972, 2576; J. R. Hanson, T. Marten and M. Siverns, *J. Chem. Soc., Perkin Trans. I* 1974, 1033.

(Received in Germany 28 August 1984)